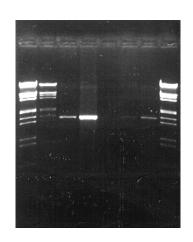
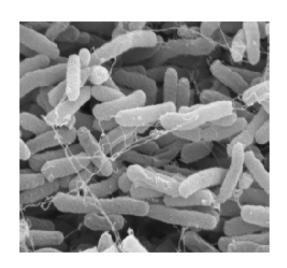


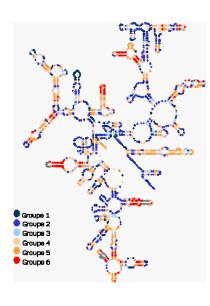
Microbiote intestinal



Antonia SUAU-PERNET



25 Avril 2018



Antonia SUAU-PERNET

Professeur des Universités

Microbiologiste moléculaire

le cnam



Enseignements au Cnam à Paris Biologie moléculaire Biologie cellulaire Microbiologie







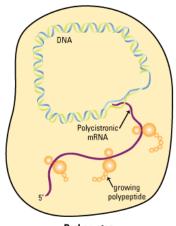


Recherche dans l'unité TIMC-IMAG, dans l'équipe GEM (Génomique et Évolution des Microorganismes) à Grenoble

Evolution d'*Escherichia coli* Microbiote intestinal

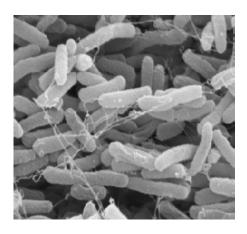
Bactérie = organisme unicellulaire microscopique

Pas observable à l'œil nu



Prokaryotes

Microscope



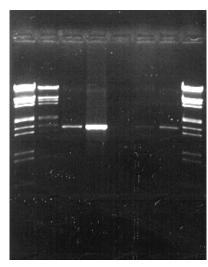
Zoom

Culture



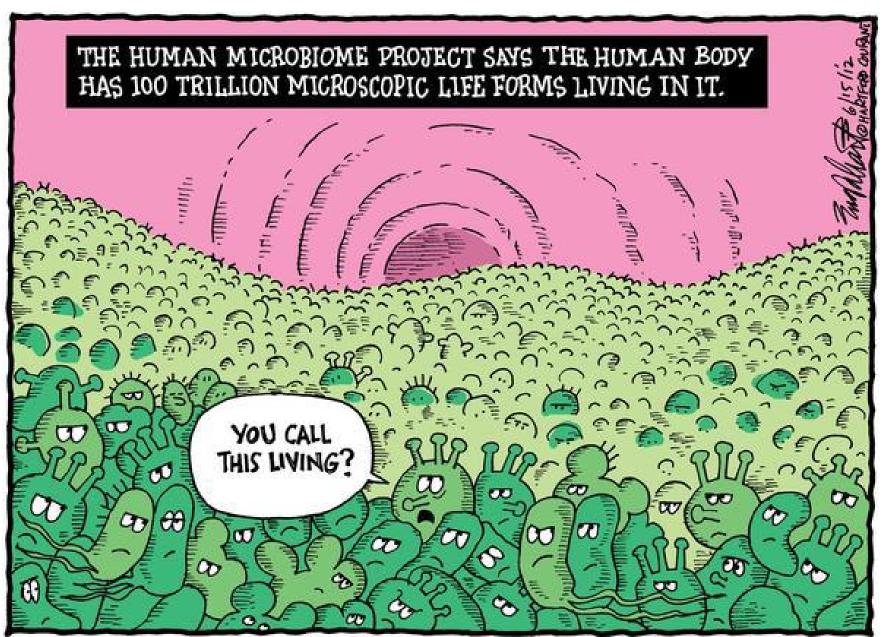
Multiplication

Outils moléculaires

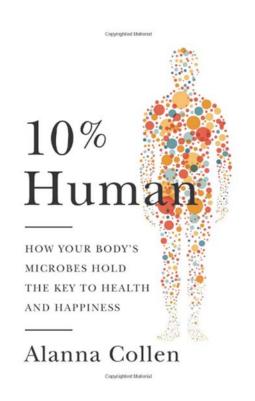


Amplification

Microbiote intestinal humain

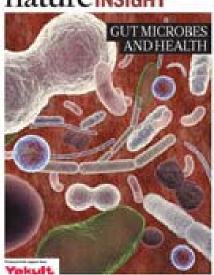


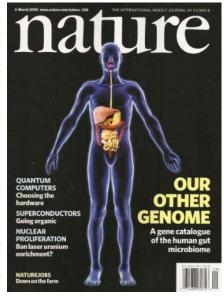




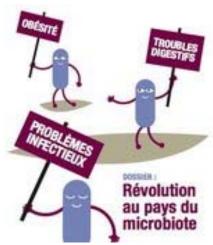






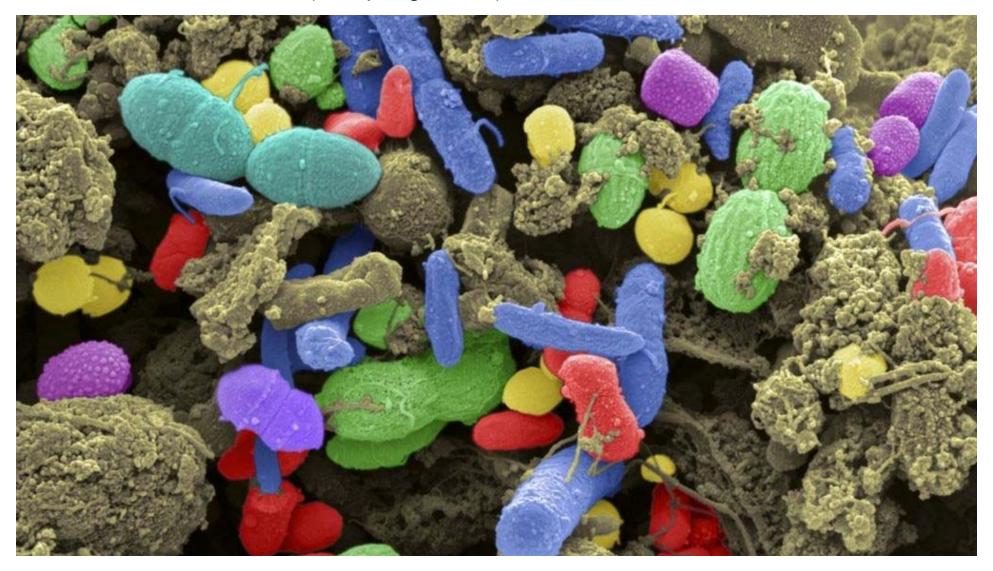






http://www.kinesport.info/Echos-de-la-micronutrition-Revolution-au-pays-du-microbiote_a1029.html

Microbiote intestinal Des milliards de bactéries (10¹² par gramme)



Microbiologiste moléculaire : étude de l'ADN des bactéries

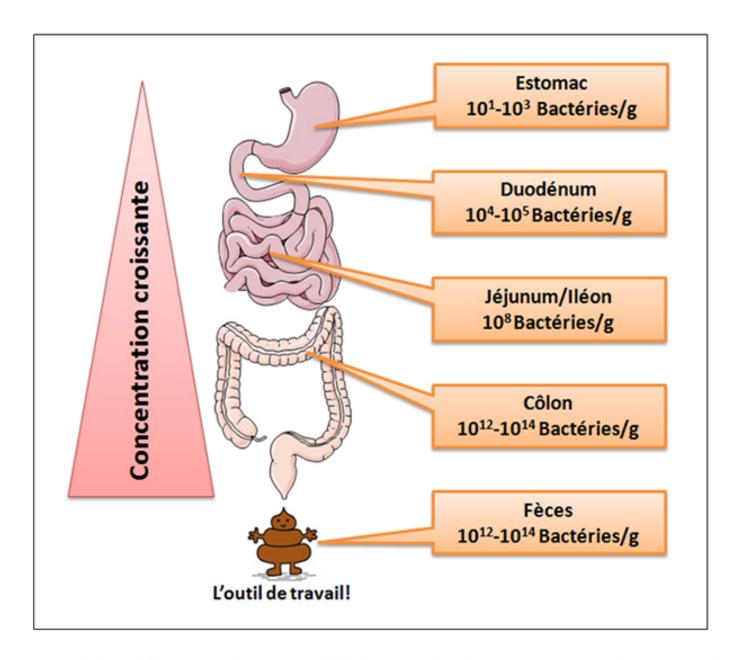
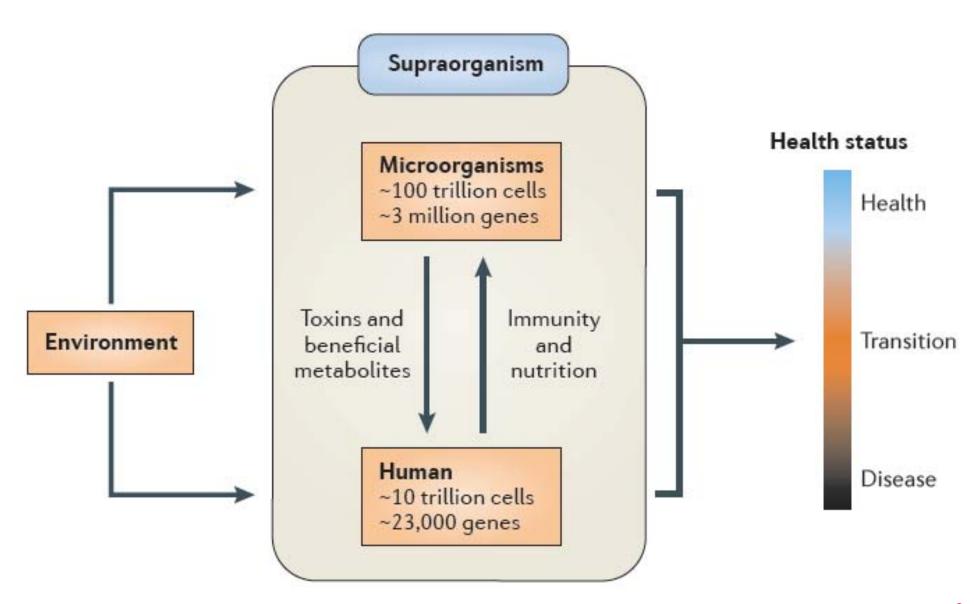


Figure 1: Répartition de la quantité de bactéries le long du tractus digestif

The gut microbiota and obesity: from correlation to causality

Liping Zhao



Trillion: 10¹²

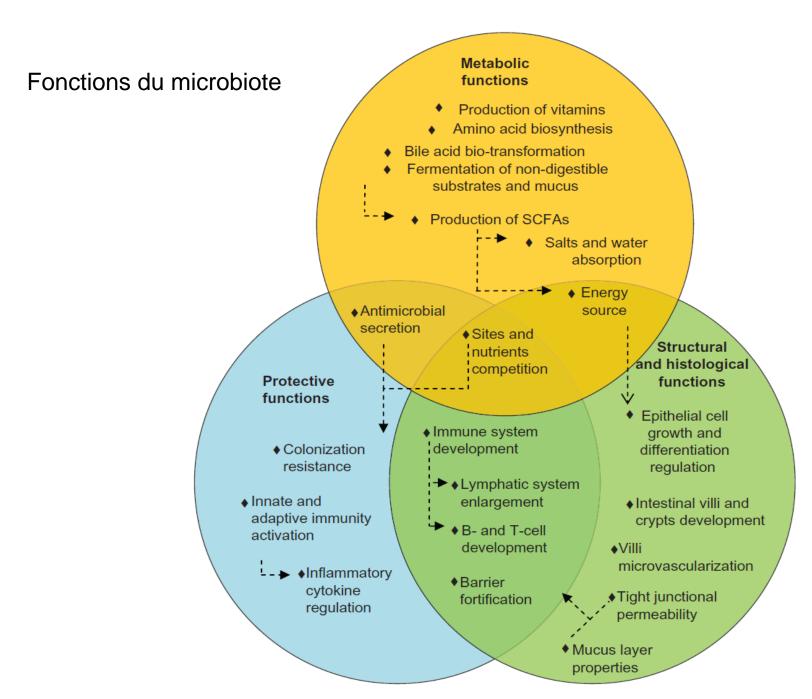
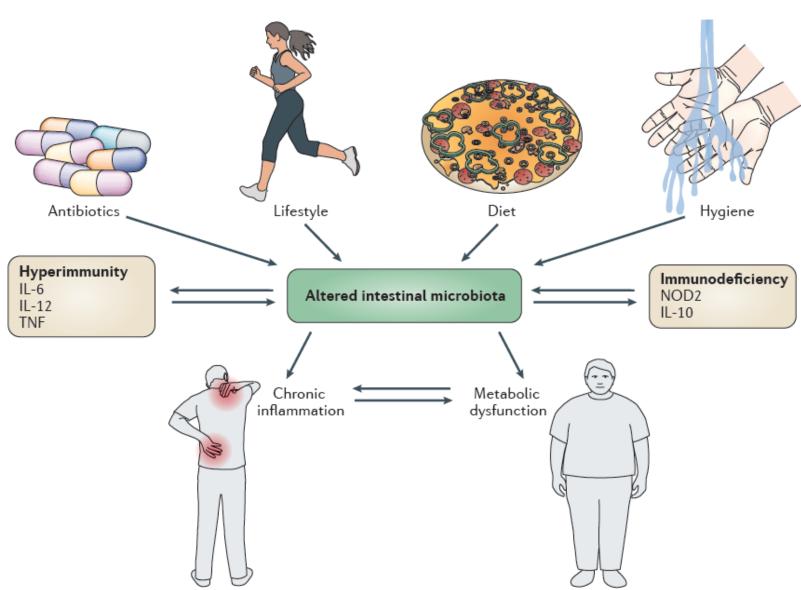


Figure I Main beneficial functions of the human gut microbiota. Circles represent the three principal classes of functions performed by the bacteria that inhabit the gut. Arrows represent causal relationships.

Abbreviation: SCFA, short chain fatty acid.

The gut microbiota — masters of host development and physiology

Felix Sommer^{1,2} and Fredrik Bäckhed^{1,2,3}



Culture

Human Fecal Flora: The Normal Flora of 20 Japanese-Hawaiians

W. E. C. MOORE AND LILLIAN V. HOLDEMAN

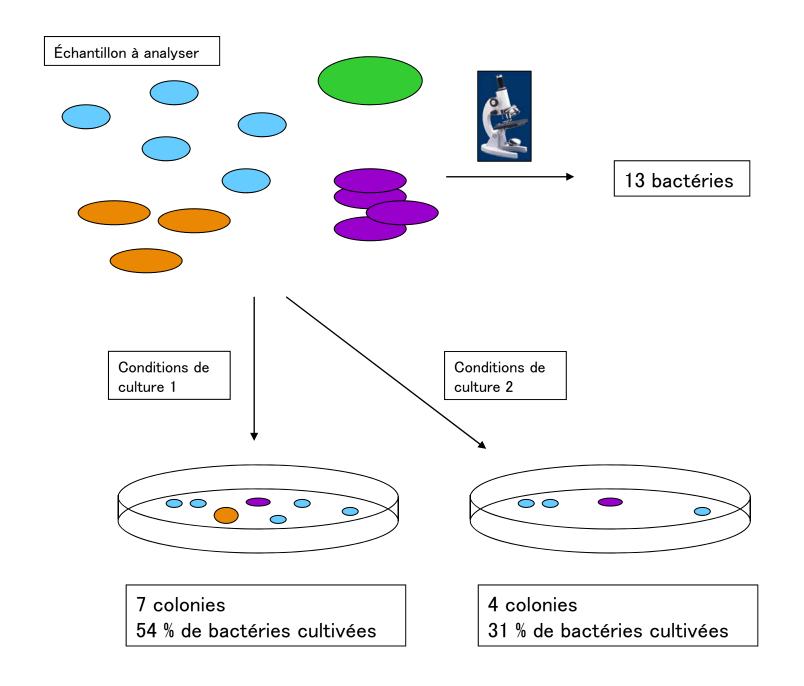
Anaerobe Laboratory, Virginia Polytechnic Institute and State University, Blacksburg, Virginia 24061

Received for publication 21 January 1974

There were 113 different kinds of organisms detected among the 1,147 isolates examined from the 20 people. In Table 3, the kinds of organisms are listed in order of frequency of their occurrence in this human population. The 113 kinds account for 94% of the viable cells (a 94% coverage) of the fecal flora of these individuals as a population.

Preliminary statistical analyses of these and additional data by I. J. Good (personal communication) indicate that the total number of different kinds of bacteria in the intestinal tract at any one time probably exceeds 400 or 500 species, but most of these are represented by less than 10⁸ cells per g of feces (less than 1/1,000 of the bacterial population).

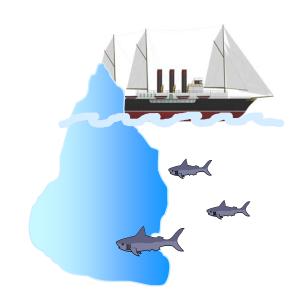
			•
Characteristic	Eubacterium - N [bals] *	Eubacterium - N-1 [bl(a)]	Eubacterium - AQ [Als2]
No. strains	4	6	1
Amygdalin Arabinose Cellobiose Dextrin Erythritol	: -	w - - - -	- - -
Esculin pH Esculin hyd. Fructose Galactose Glucose	- wa w w	- - -w - w-	_ _ _w _ a
Glycerol Glycogen Inositol Inulin Lactose	:	- -	_ _ _
Maltose Mannitol Mannose Melezitose Melibiose	v	- - -	=
Raffinose Rhamnose Ribose Salicin Sorbitol	- -a - -w -	- - -	_ a a
Sorbose Starch pH Starch hyd. Sucrose Trehalose Xylose	· · · · · · · · · · · · · · · · · · ·	w- 	1
Gelatin dig. Milk Indole EYA react. Hemolysis	- - -	- - -	=======================================
H ₂ S Gas Hydrogen PY-growth PY-CHO gr. PYG-bile gr.	1 4 -,1 2	- - 1 4 3	- 4 4 2 4 2,3
Pyruvate Gluconate Motility	(A) - -	=	<u> </u>
Morphology 10 microns	一方子	18 18 18 18 18 18 18 18 18 18 18 18 18 1	3



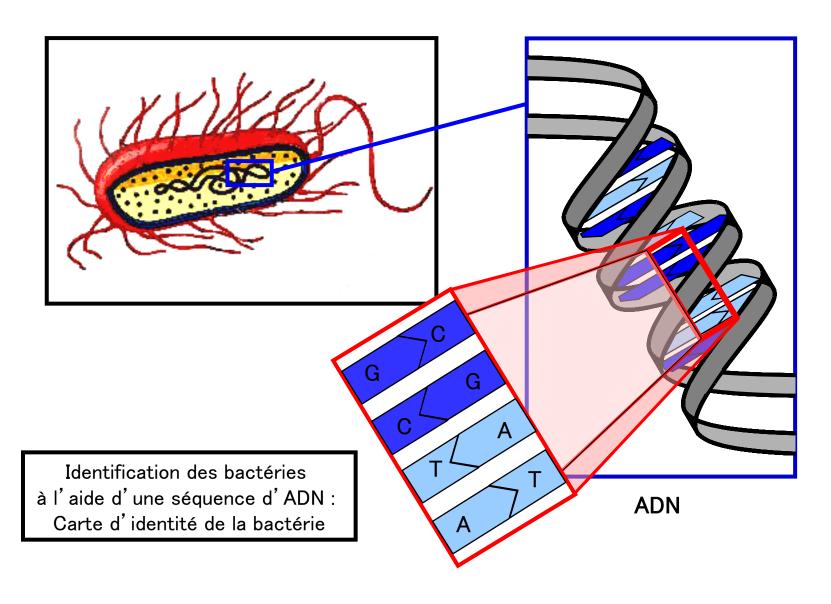
Variabilité des pourcentages de bactéries cultivées

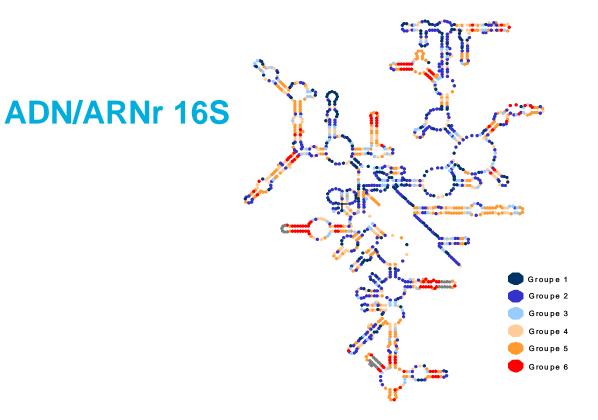
Une large proportion du microbiote intestinal adulte demeure "non-cultivée"

Références	Fraction cultivée	
Moore et Holdeman, 1974	37 - 464 %	
Langendijk <i>et al.,</i> 1995	14 - 37%	
Wilson et Blitchington, 1996	58%	
Suau <i>et al.,</i> 1999	20 - 30%	
Hayashi <i>et al.,</i> 2002	38 %	



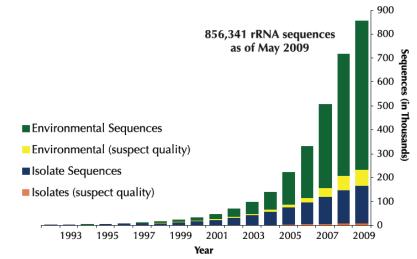
Identification moléculaire des bactéries





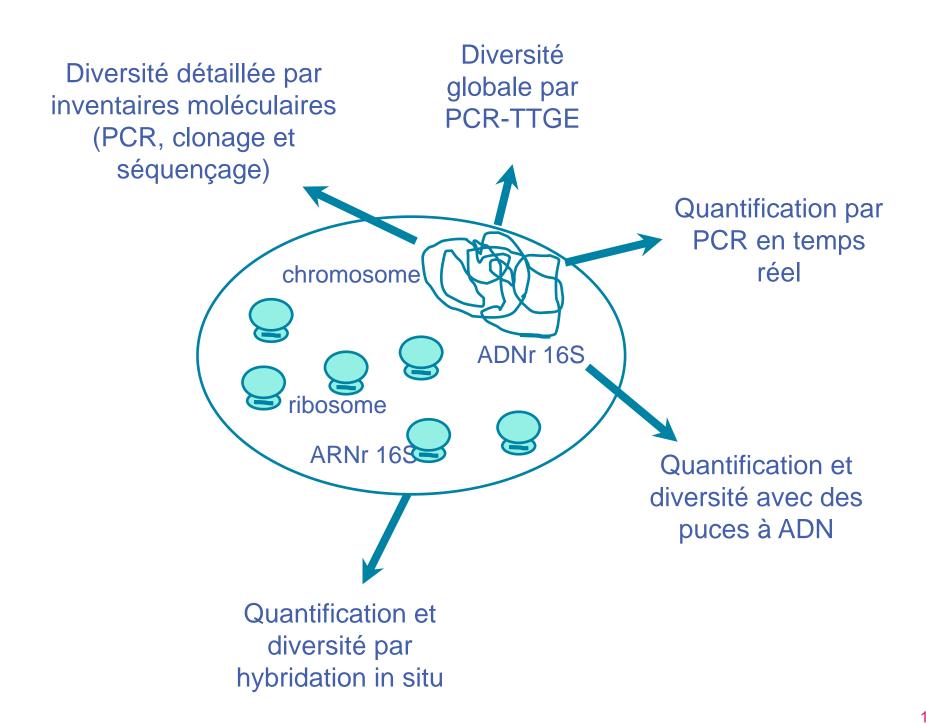
Variabilité de l'ARNr 16S

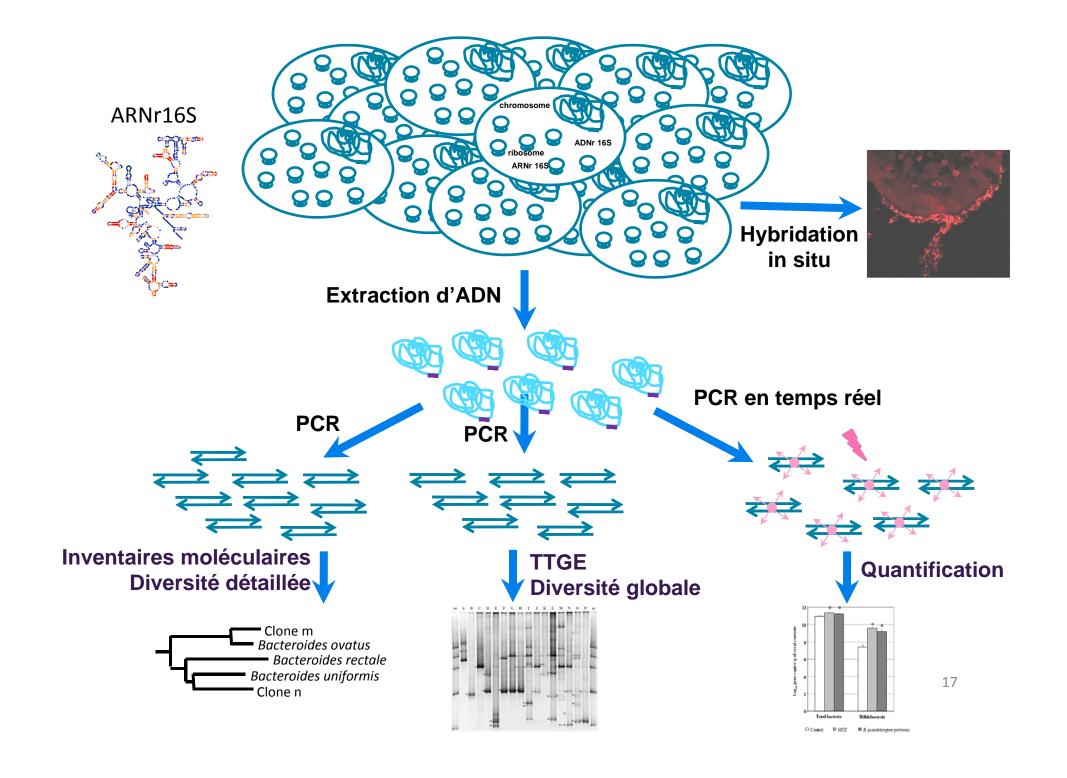
Les nucléotides sont divisés en six groupes de variabilité croissante (1 à 6).
Les nucléotides présents chez *E. coli* mais absents chez 75 % des autres micro-organismes sont en gris.
Copyright Yves Van de Peer.

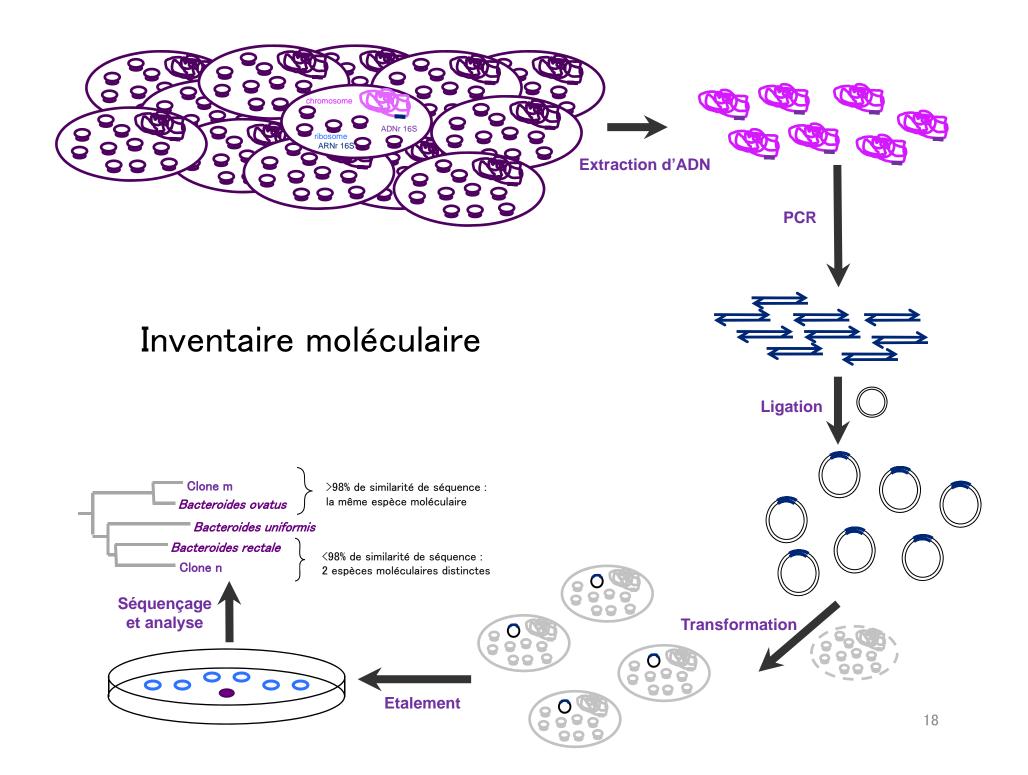


Poster ASM, 2009

RDP Release 11, Update 5 September 30, 2016 3,356,809 16S rRNAs 125,525 Fungal 28S rRNAs



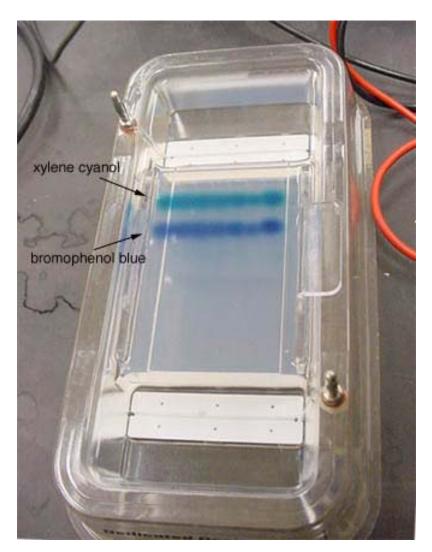




Electrophorèse en gel d'agarose







PCR

Réaction de polymérisation en chaîne Polymerase chain reaction

Réplication d'ADN in vitro

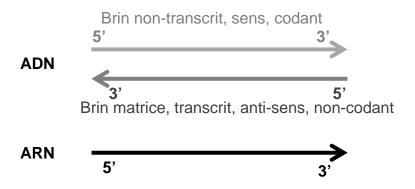
ADN matrice
Polymérase
Amorces (primers)
dNTP
MgCl₂
Tampon 10X



thermocycleur



microtube

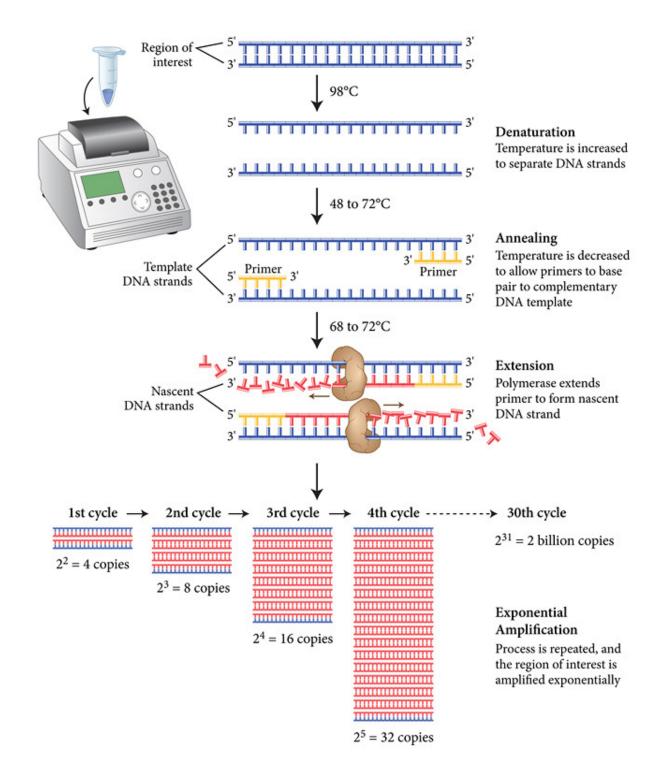




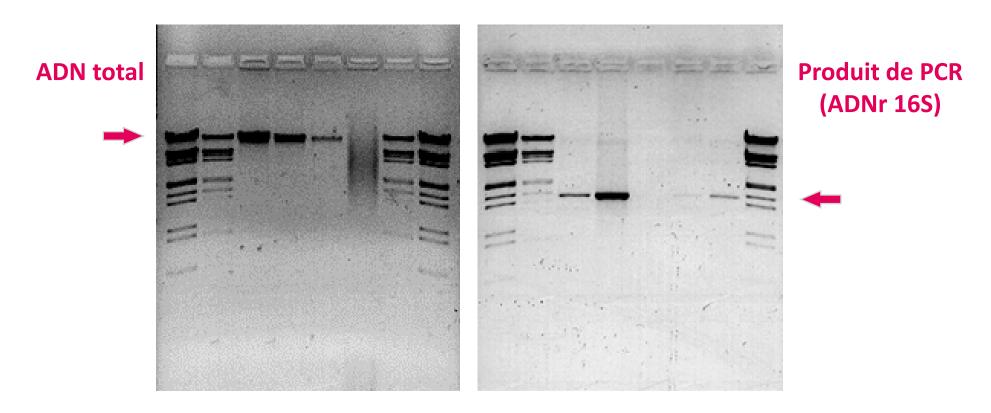
Amorce forward identique à l'extrémité 5' du brin sens s'hybride au brin matrice permet la synthèse d'un brin sens

Amorce reverse identique à l'extrémité 5' du brin matrice s'hybride au brin sens permet la synthèse d'un brin matrice

Schéma de principe de la PCR



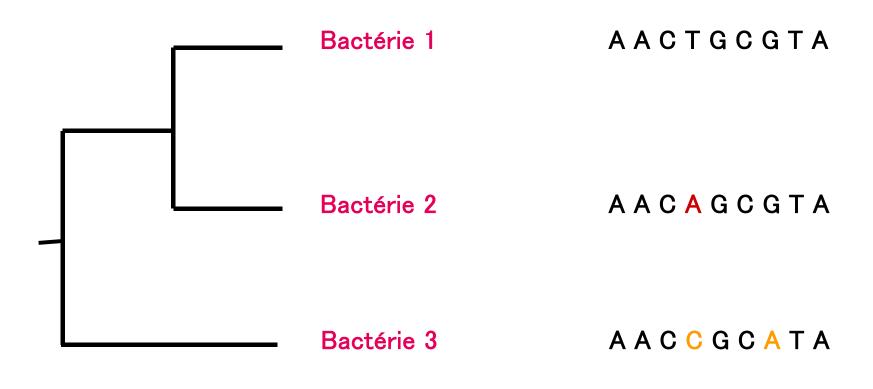
Electrophorèses en gel d'agarose







Principe de construction d'un arbre phylogénétique



Arbre phylogénétique

Alignement de séquences



Qu'est ce que l'espèce moléculaire ?

Clones et souches bactériennes sont groupés dans une espèce moléculaire ou OTU (Operational Taxonomic Unit) quand la similarité entre leurs séquences d'ADNr 16S est supérieure à 98%.

Séquençage

Direct Analysis of Genes Encoding 16S rRNA from Complex Communities Reveals Many Novel Molecular Species within the Human Gut

ANTONIA SUAU, ^{1,2}* RÉGIS BONNET, ² MALÈNE SUTREN, ¹ JEAN-JACQUES GODON, ³ GLENN R. GIBSON, ² MATTHEW D. COLLINS, ² AND JOEL DORÉ ¹

The human intestinal tract harbors a complex microbial ecosystem which plays a key role in nutrition and health. Although this microbiota has been studied in great detail by culture techniques, microscopic counts on human feces suggest that 60 to 80% of the observable bacteria cannot be cultivated. Using comparative analysis of cloned 16S rRNA gene (rDNA) sequences, we have investigated the bacterial diversity (both cultivated and noncultivated bacteria) within an adult-male fecal sample. The 284 clones obtained from 10-cycle PCR were classified into 82 molecular species (at least 98% similarity). Three phylogenetic groups contained 95% of the clones: the *Bacteroides* group, the *Clostridium coccoides* group, and the *Clostridium leptum* subgroup. The remaining clones were distributed among a variety of phylogenetic clusters. Only 24% of the molecular species recovered corresponded to described organisms (those whose sequences were available in public databases), and all of these were established members of the dominant human fecal flora (e.g., *Bacteroides thetaiotaomicron*, *Fusobacterium prausnitzii*, and *Eubacterium rectale*). However, the majority of generated rDNA sequences (76%) did not correspond to known organisms and clearly derived from hitherto unknown species within this human gut microflora.

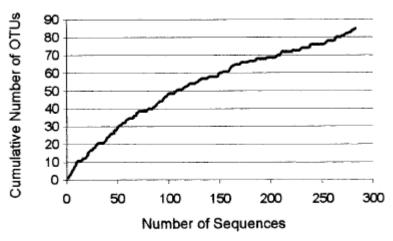


FIG. 4. Estimation of the biodiversity which was obtained by direct community analysis of a fecal sample. The cumulative number of OTUs is given as a function of the number of clones sequenced. Clones were randomly used.

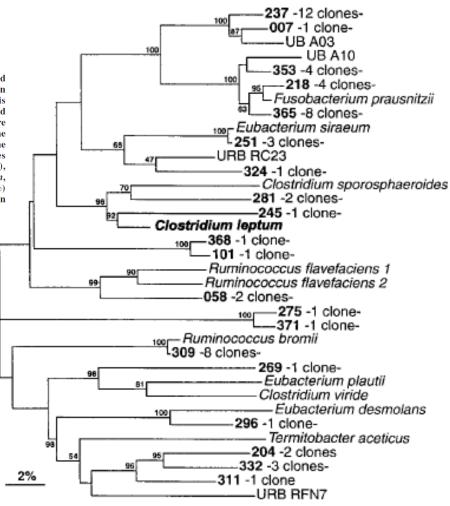
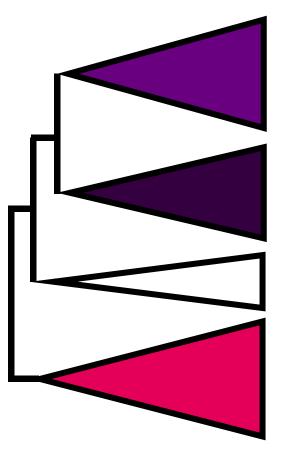


FIG. 3. Phylogenetic tree derived from partial 16S rDNA sequence data for members of the *Clostridium leptum* subgroup. Bar represents 2% sequence divergence. Designations of clones and the key organism used to name the group are in boldface type. The tree was constructed with the SIMILARITY and NEIGHBOR programs. Bootstrap values are based on 500 replications.

Groupes phylogénétiques de la microflore fécale humaine d'un adulte sain



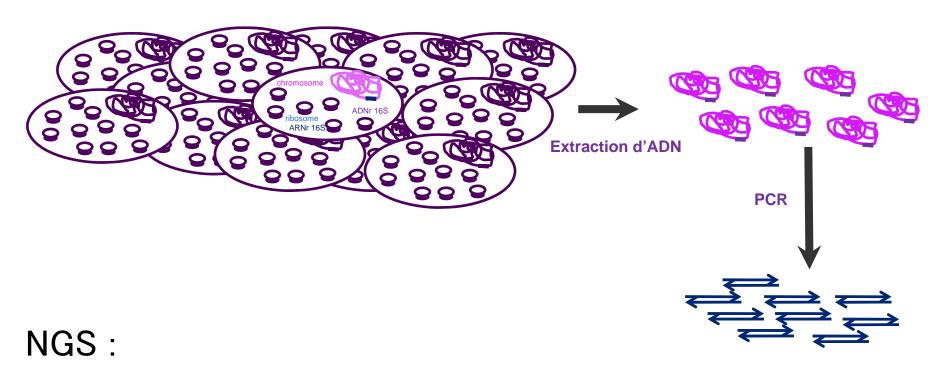
Famille *Ruminococcaceae*43% clones, 31 "espèces", 8 décrites*

Famille *Lachnospiraceae*21% clones, 20 "espèces", 3 décrites*

Genre Bifidobacterium aucun clone!

Phylum *Bacteroidetes* 31% clones, 20 "espèces", 8 décrites *

^{* &}gt; 98% similarité avec 1 séquence connue



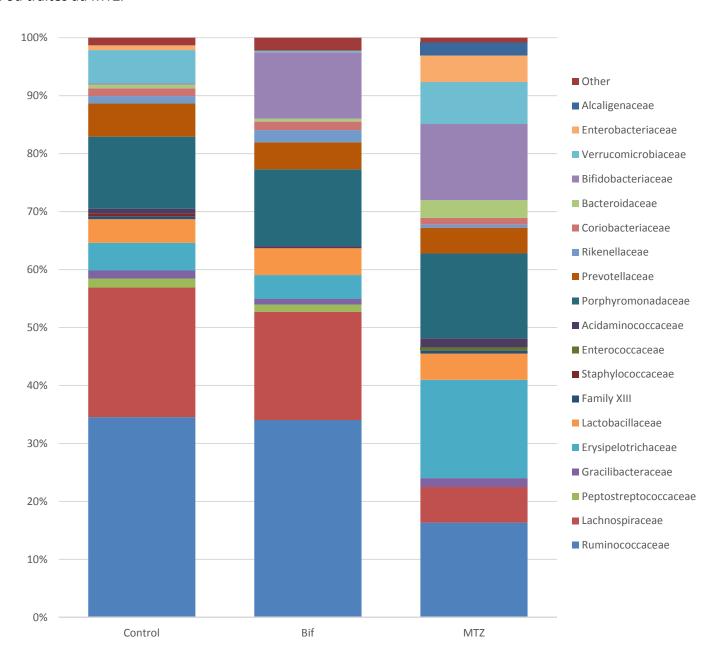
Séquençage de nouvelle génération

Pas besoin de clonage Beaucoup plus de séquences Moins cher



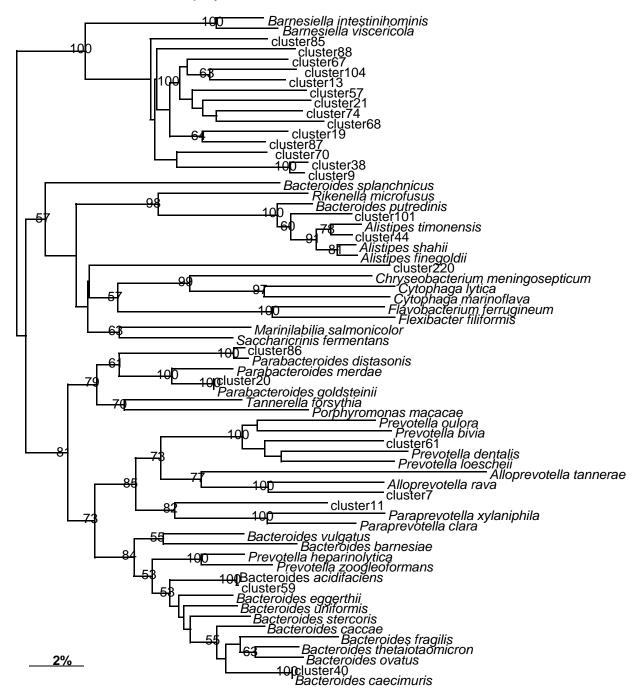
Structure de la communauté bactérienne au niveau des familles

L'abondance est présentée en termes de pourcentage de séquences bactériennes dans le microbiote des rats témoin, gavés avec la souche de bifidobactérie ou traités au MTZ.

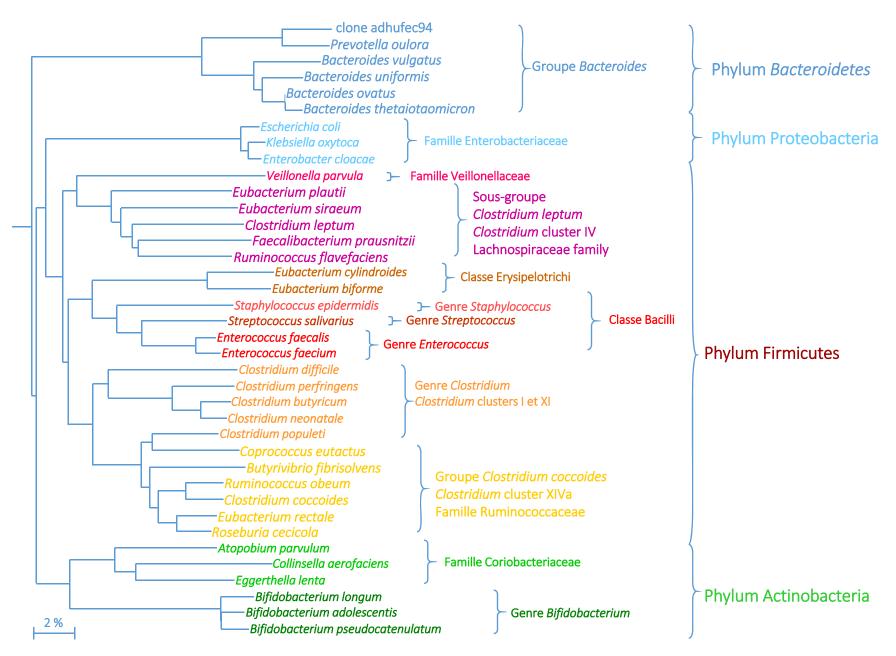


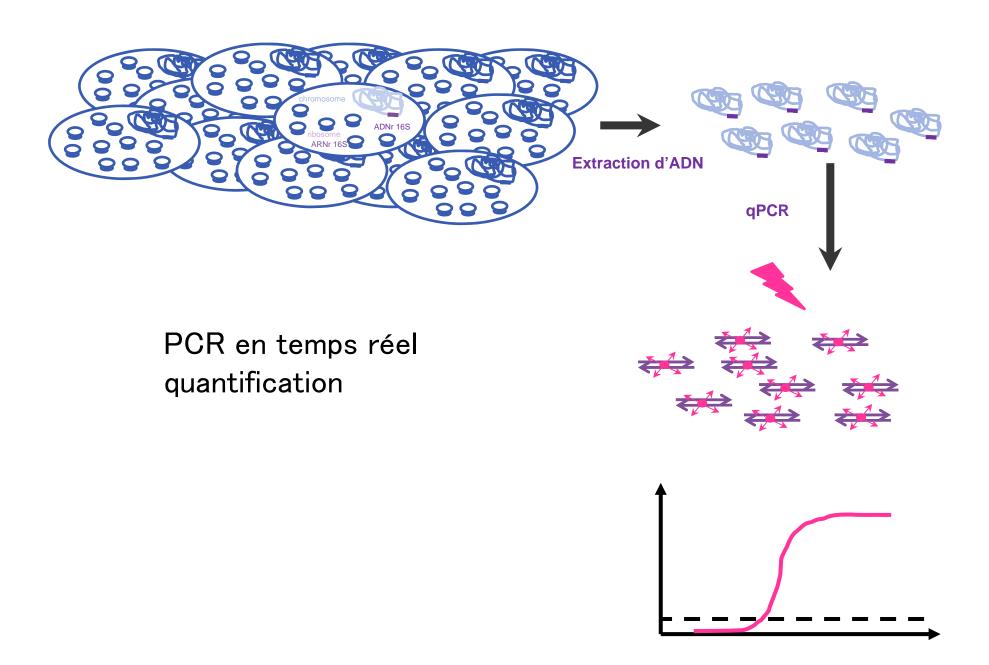
30

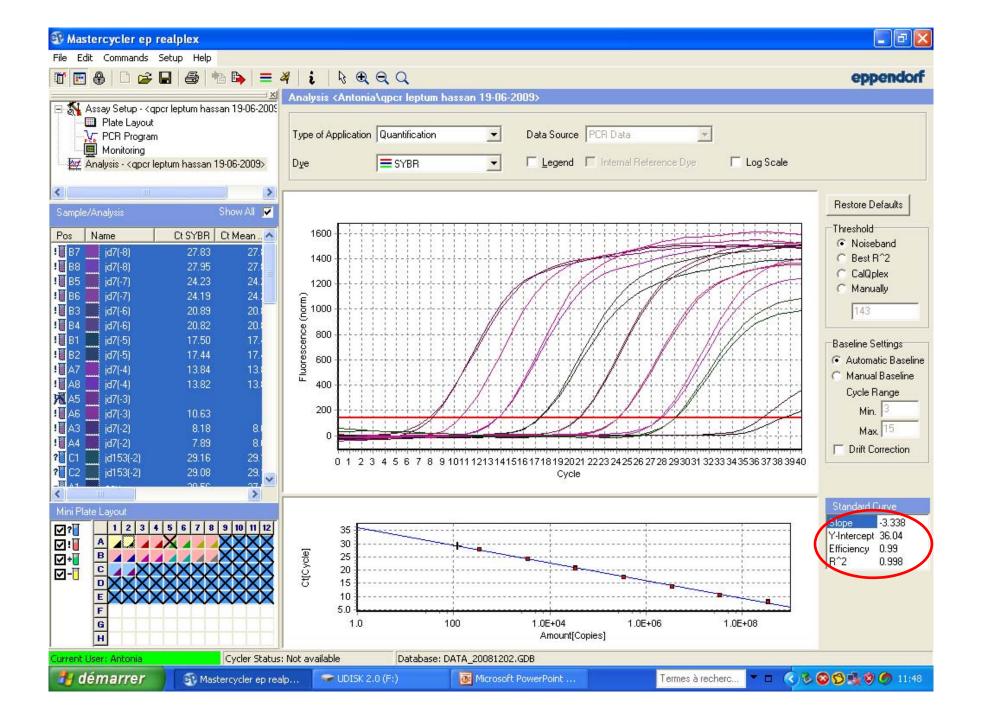
Phylogenetic tree of the Bacteroidetes phylum for the most abundant OTUs



Microbiote intestinal analysé par la biologie moléculaire

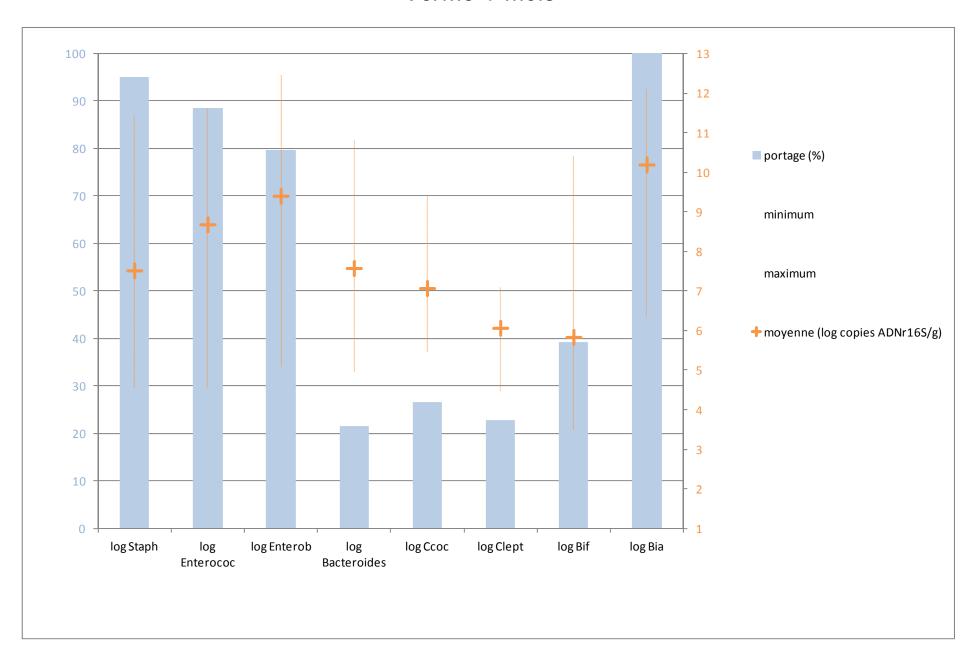


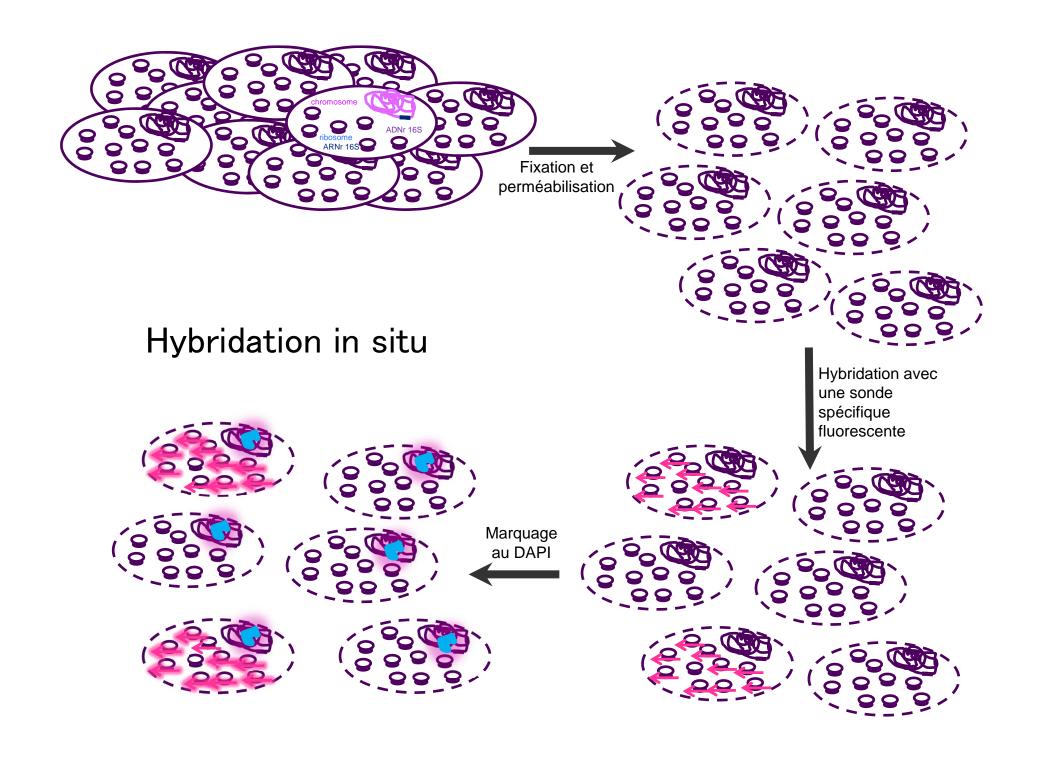




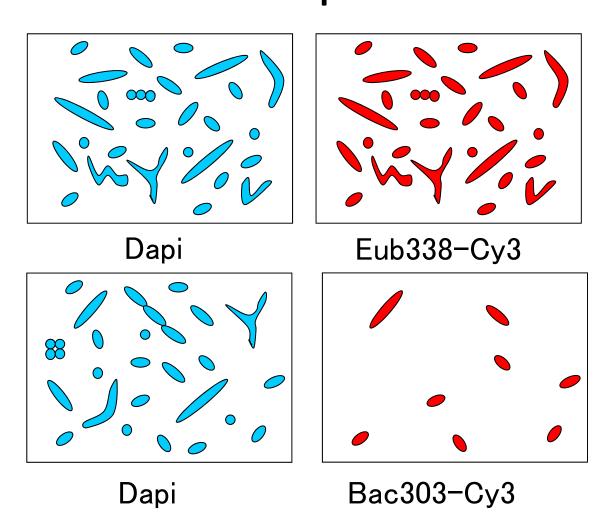


Projet Prémaflora Terme 1 mois





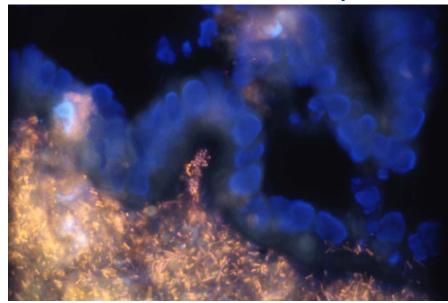
Hybridation In Situ couplée à l'analyse d'images -Principe-



Hybridation in situ sur caecum de souris avec la sonde générale Bacteria

Coloration Dapi et fluorescence Eub 338-Cy3

Fluorescence Eub 338-Cy3

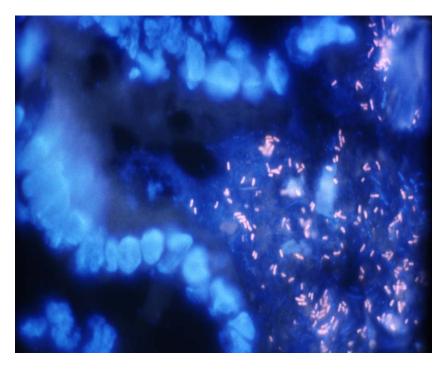




Hybridation in situ sur caecum de souris avec la sonde ciblant les Bifidobactéries

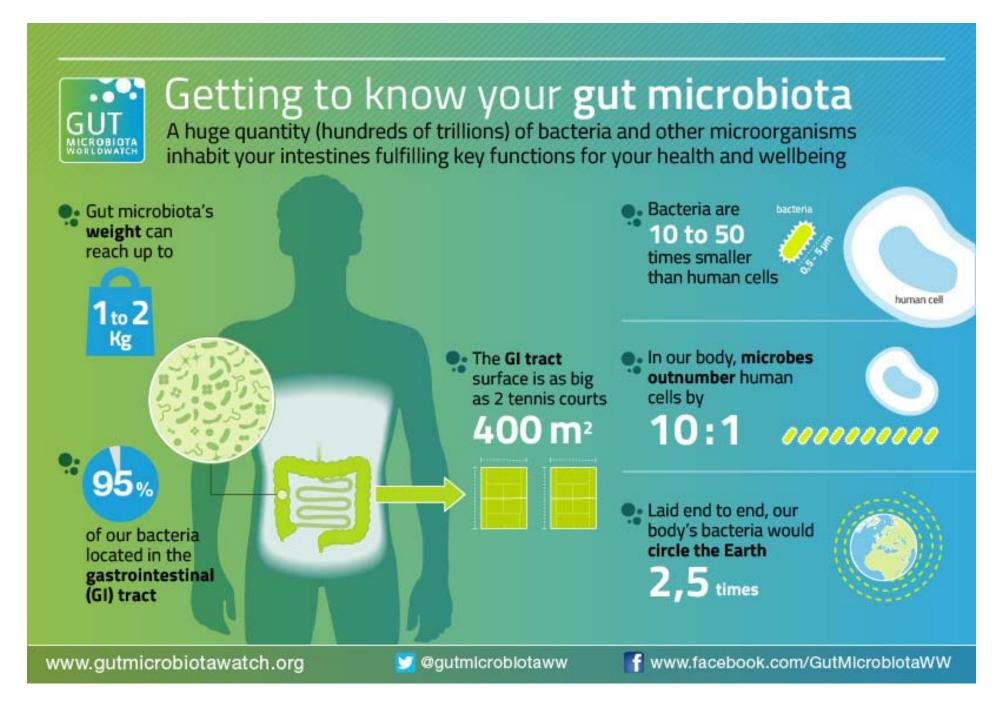
Observation coloration Dapi et fluorescence Bif 164-Cy3

Fluorescence Bif 164-Cy3





Côlon d'un rat du Côlon d'un rat groupe contrôle traité au MTZ Epaisseur de la couche de mucus (coloration bleu alcian / acide périodique/ réactif de Schiff) Hybridation in situ avec la sonde Bacteria CY3 Marquage au DAPI



Implantation du microbiote

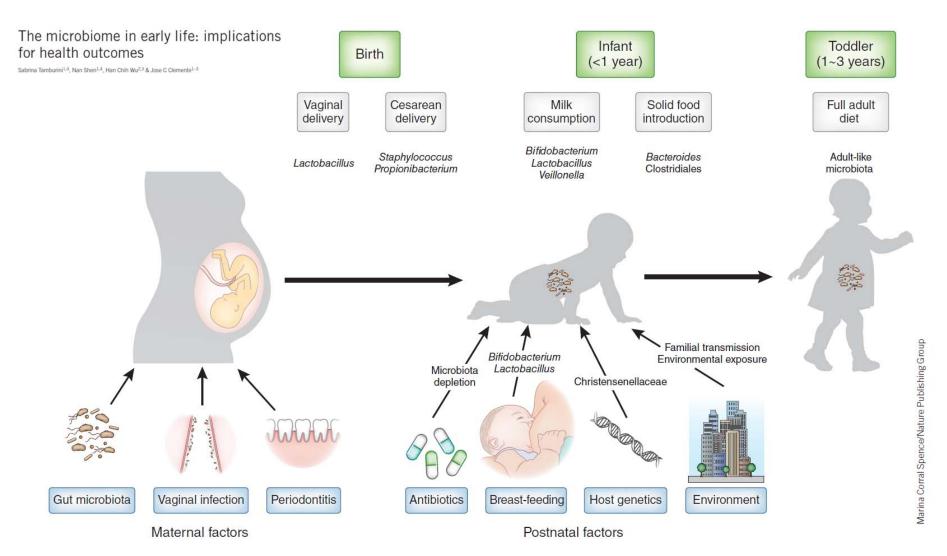


Figure 1 Factors shaping the neonatal microbiome. Maternal vaginal infections or periodontitis can result in bacteria invading the uterine environment. Gut and oral microbiota could be transported through the bloodstream from the mother to the fetus. Delivery mode shapes the initial bacterial inoculum of the newborn. Postnatal factors such as antibiotic use, diet (such as breast-feeding versus formula, and introduction of solid food), genetics of the infant and environmental exposure further configure the microbiome during early life. As diet diversifies with age, the microbiome gradually shifts toward an adult-like configuration, which is usually reached by age 3. Bacteria associated with the different processes are indicated.

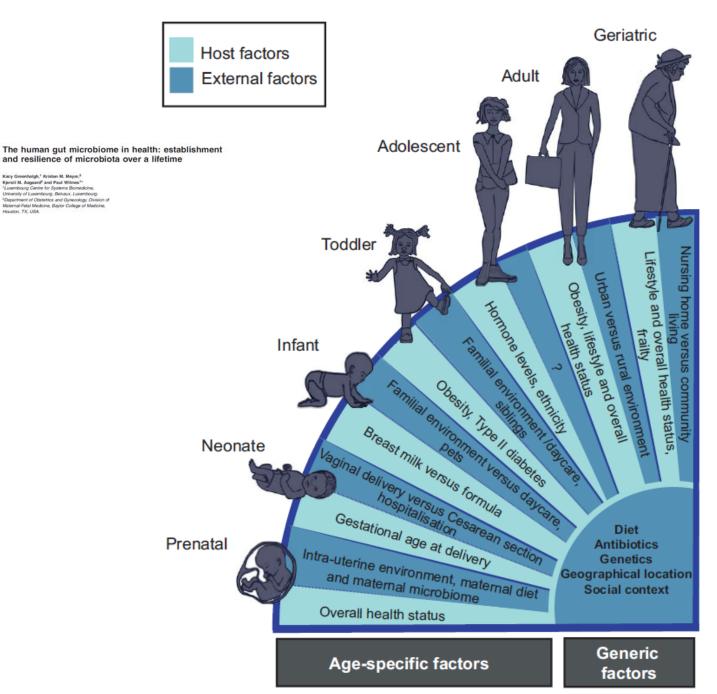
The human gut microbiome in health: establishment and resilience of microbiota over a lifetime

Kacy Greenhalgh,¹ Kristen M. Meyer,²
Kjersti M. Aagaard² and Paul Wilmes^{1*}
¹Luxembourg Centre for Systems Biomedicine,
University of Luxembourg, Belvaux, Luxembourg.
²Department of Obstetrics and Gynecology, Division of
Maternal-Fetal Medicine, Baylor College of Medicine,
Houston, TX, USA.





Interindividual variability



Kacy Greenhalgh, 1 Kristen M. Meyer, 2 Kjersti M. Aagaard and Paul Wilmes^{1,4} *Lusembourg Centrle for Systems Blomedicine, University of Lusembourg, Belviux, Lusembourg, *Obpartment of Obstetics and Operacology, Division of Material-Patal Medicine, Baylor College of Medicine, Houston, TX, USA

Fig. 2. Factors which influence the gastrointestinal tract microbiome according to different life stages.

Modifications du microbiote intestinal

Emerging Technologies for Gut Microbiome Research

Jason W. Arnold, ¹ Jeffrey Roach, ² and M. Andrea Azcarate-Peril ^{1,*}

Modulator Type	Example	Impact on Host	Duration/rate	Effects on the Microbiota
Prebiotics	β,1-4 Galacto- oligosaccharide (GOS)	Beneficial	Short-term/ intermediate	Promotes growth of Bifidobacterium and Lactobacillus [121] Inhibits growth of Clostridium [122] Increases recovery rate of microbiota post-antibiotic treatment [123]
Probiotics	Lactobacillus rhamnosus GG	Beneficial	Short-term/ intermediate	Inhibits growth/colonization of pathogenic microbes [90,124] Promotes growth of Bifidiobacterium sp. [125] Modulates host gene expression [90]
Bacteriophages	933 W coliphage	Detrimental	Short-term/rapid	Modify and/or eradicate populations of commensal <i>Escherichia coli</i> [126] Transmits endotoxin genes to bacteria within community [127]

Antibiotics/drugs	Chemotherapy	Beneficial or detrimental	Short-term/rapid	Culling of microbes to free niche space [128]
Host immune response	Toll-like receptor- mediated gene expression	Generally beneficial	Long-term/rapid	Inhibits colonization of certain microbes Eliminates invading pathogens from the population [46]
Diet	High-fat vs. low-fat diets	Beneficial or detrimental	Long-term/slow	Vitamin supplements impact transcription and microbial content [82,129] High-fat diets promote 'unhealthy' microbiota [80]
Transplantation	Fecal transplant	Mostly beneficial	Long-term/ immediate	Transplantation of healthy microbiota can eliminate Clostridium difficile infection [97]
Pathogenic bacteria	Salmonella sp.	Detrimental	Short-term/ intermediate	Outcompetes other microbes within the microbial community, reducing community diversity [130]

